



Teaching Guide				
Identifying Data			2016/17	
Subject (*)	Bioquímica: Bioquímica II	Code	610G02012	
Study programme	Grao en Bioloxía			
Descriptors				
Cycle	Period	Year	Type	Credits
Graduate	1st four-month period	Second	FB	6
Language	Spanish			
Teaching method	Face-to-face			
Prerequisites				
Department	Bioloxía Celular e Molecular			
Coordinador	Gonzalez Siso, Maria Isabel	E-mail	isabel.gsiso@udc.es	
Lecturers	Cerdan Villanueva, Maria Esperanza Freire Picos, María Ángeles Gonzalez Siso, Maria Isabel Lamas Maceiras, Mónica Varela Eirín, Marta Vizoso Vázquez, Ángel José	E-mail	esper.cerdan@udc.es maria.freirep@udc.es isabel.gsiso@udc.es monica.lamas@udc.es marta.varelae@udc.es a.vizoso@udc.es	
Web	ciencias.udc.es/bcm			
General description	The subject contents provide to the student the basic information for knowing and understanding the biochemical reactions, enzyme catalysis and metabolism. Studying in the second year of Biology degree will allow the student to achieve the basic knowledge to understand the molecular mechanisms governing many responses in the live organisms.			

Study programme competences / results	
Code	Study programme competences / results
A8	Illar, analizar e identificar biomoléculas.
A10	Avaliar actividades metabólicas.
A26	Deseñar experimentos, obter información e interpretar os resultados.
A29	Impartir coñecementos de Bioloxía.
A30	Manexar adecuadamente instrumentación científica.
A31	Desenvolverse con seguridade nun laboratorio.
B1	Aprender a aprender.
B2	Resolver problemas de forma efectiva.
B3	Aplicar un pensamento crítico, lóxico e creativo.
B4	Traballar de forma autónoma con iniciativa.
B5	Traballar en colaboración.
B7	Comunicarse de maneira efectiva nunha contorna de traballo.
B10	Exercer a crítica científica.
B11	Debater en público.
B13	Comportarse con ética e responsabilidade social como cidadán e como profesional.

Learning outcomes	
Learning outcomes	Study programme competences / results



Understand and describe the mechanisms by which the Enzymes act in biological catalysis. To design, combining and using the methodology of the practical course and theoretical classes, systems to purify and analyse enzymes. To know the importance of the pathways to obtain energy in the biological systems to maintain life. To know the main metabolic pathways in the cell and its regulation. Develop the capability to relate the different metabolic pathways.	A8	B1
	A10	B2
	A26	B3
	A29	B4
	A30	B5
	A31	B7
		B10
		B11
		B13

Contents	
Topic	Sub-topic
1. Biological Enzymes as catalysts.	Structural features that give them advantages over chemical catalysts. Reaction mechanisms. Antibodies as catalysts. Ribozymes.
2. Kinetics of chemical reactions.	Monosubstrate reactions and Michaelis-Menten kinetics Transformation of the Michaelis- Menten equation. Bisubstrate reaction kinetics. Irreversible inhibitors; binding, examples and applications. Reversible Inhibition: types of inhibition. Kinetics in the presence of inhibitors.
3. Regulation of enzymatic activity.	Importance of regulation in metabolism. Allosteric enzymes. Covalent modification. Isoenzymes. Zymogens or proenzymes.
4. Methodology for determination of enzyme activities.	Direct and indirect assays. Purification of enzymes: specific activity, yield and purification factor. Importance and current applications of enzymology.
5. Introduction to Metabolism.	Anabolic and catabolic pathways. Compartmentalization. Need for coordination and interaction between the different routes, and variability among species. Levels of obtaining energy. Methodology for the study of metabolic pathways. Levels of study.
6. Transport of metabolites across cell membranes.	Types of transport depending on the energy sources. Structural data. Examples with specific metabolites
7. Obtaining chemical energy.	Oxidation reduction in energy production. Coenzymes involved. Generation of ATP: substrate-level phosphorylation, oxidative phosphorylation and photosynthetic phosphorylation and energy production systems. Detailed study of oxidative phosphorylation and photosynthetic phosphorylation.
8: Glycolysis and catabolism of hexoses.	Location of the routes. Stages and pathway regulation. Fermentations. Relationship with the pentose phosphate pathway.
9: TCA cycle	Location of the route. Conversion of pyruvate to acetyl-CoA. Study of the pyruvate dehydrogenase complex and interaction with other routes. Anaplerotic routes, importance of mitochondrial shuttles and balances.
10. Gluconeogenesis.	Definition and localization, metabolic need for this route. Gluconeogenesis from: pyruvate, lactate, amino acids and triglycerides. Glyoxylate cycle.
11. "Dark Phase" of photosynthesis. Relationship with gluconeogenesis.	The Calvin cycle. Photorespiration. Regulation. The C4 pathway of tropical plants. The crassulacean acid metabolism. Sucrose metabolism and starch.
12. Glycogen metabolism.	The reserve polysaccharide glycogen. Biosynthesis and degradation of muscle and liver glycogen. Regulation. The role of the liver in the maintenance of blood glucose. Congenital anomalies of glycogen metabolism
13. Lipid Metabolism.	Lipid catabolism: lipolysis, beta-oxidation. Biosynthesis of fatty acids, triglycerides, membrane lipids and steroids. Regulation of lipid metabolism. Metabolism of ketone bodies.



14. Metabolism of amino acids.	Digestion and degradation of intracellular proteins. Nitrogen removal of amino acids: transamination, deamination. Urea cycle. Ammonia transport to the liver. Fate of the carbon skeleton of amino acids. Amino acid biosynthesis: origin of nitrogen and carbon skeleton. regulation
15. Derivatives of amino acids.	Amino acid precursor functions: Amines with biological activity, glutathione, porphyrins. Metabolism of purine and pyrimidine nucleotides. regulation
16. Integration of metabolism.	Metabolic profiling of major organs. Key connections between routes: glucose-6-phosphate, pyruvate and acetyl CoA. Metabolic adaptations to stress. Fasting, exercise.
17. Hormonal regulation of metabolism.	Hormones as chemical messengers. Second messengers. Metabolic targets of hormone action. Hormone receptors. Adenylate cyclase system. Phospholipase system. Receptor dimerization

Planning				
Methodologies / tests	Competencies / Results	Teaching hours (in-person & virtual)	Student?s personal work hours	Total hours
Guest lecture / keynote speech	A10 B1 B3 B4 B7 B10 B11 B13	24	60	84
Problem solving	A10 A29 B1 B2 B3 B4 B5 B7 B10 B11 B13	8	16	24
Laboratory practice	A8 A26 A30 A31 B1 B2 B3 B4 B5 B7 B10 B13	15	22.5	37.5
Mixed objective/subjective test	A8 A10 A26 B2 B3 B7 B13	2	0	2
Personalized attention		2.5	0	2.5

(\*)The information in the planning table is for guidance only and does not take into account the heterogeneity of the students.

Methodologies	
Methodologies	Description
Guest lecture / keynote speech	Oral presentation supplemented with the use of media and the introduction of some questions to the students, in order to impart knowledge and facilitate learning. The master class is also known as "conference" , " expository method " or " maxistral lesson." The latter method is usually reserved for a special kind of lesson taught by a teacher in special occasions, containing original preparation involved and based on the use of the word and images as a means of transmission of information to the audience.
Problem solving	Technique by which you have to solve a specific problem situation, from the previous theoretical resources, and may have more than one possible solution.
Laboratory practice	Methodology that allows students actually learn -as through conducting practical activities, such as demonstrations exercises, experiments and research.
Mixed objective/subjective test	Exam that integrates objective test, short questions and resolution of cases and issues.

Personalized attention	
Methodologies	Description



Guest lecture / keynote speech	The personalized attention will be just along the course and at the request of the student.
Problem solving	The way of working to acquire skills and prepare the exams, the orientation of problem solving and interpretation of results of practical work, as well as any other questions that arise from the student shall be guided by this personal attention.
Laboratory practice	Students with part-time dedication or waiver of assistance should contact the teachers of the subject in the early going to establish a schedule of activities to acquire and conveniently evaluate the skills of matter.
Mixed objective/subjective test	

Assessment			
Methodologies	Competencies / Results	Description	Qualification
Problem solving	A10 A29 B1 B2 B3 B4 B5 B7 B10 B11 B13	Student's work in reduced groups and controls.	20
Laboratory practice	A8 A26 A30 A31 B1 B2 B3 B4 B5 B7 B10 B13	Participation in the clases and Exam.	15
Mixed objective/subjective test	A8 A10 A26 B2 B3 B7 B13	The knowledgements aquired by the students in both, the master clases and the problem solving will be evaluated.	65

Assessment comments
<p>Attendance at the laboratory practice is mandatory to pass the subject. Assistance may be validated, upon request, by the already completed in the immediately preceding year. If the student does not attend to laboratory work, the student must pass a test to demonstrate the practical training. In addition to attending practices, the skills acquired will be evaluated through a exam.</p> <p>Attendance and participation of students in small groups (problem solving) are valued by their work in groups, preparation of schemes and controls.</p> <p>In the January option to pass the course. the student must have at least 26 points in the mixed test of 65 total, and adding the other activities more than 50 points. If not reached 26 points in the mixed test, the other activity scores are not added (laboratory and small groups).</p> <p>For the July rating, student may kept approved parts in January and pass only the remaining parts. Alternatively, in July the student can recover 100% of the grade for the course through a practice exam (15%) and a mixed test (85%).</p> <p>For students with part-time dedication or waiver assistance, in January and July there will be a specific exam for overall assessment.</p>

Sources of information	
Basic	<ul style="list-style-type: none"> <li>- Feduchi, Blasco, Romero y Yáñez (2011). Bioquímica, conceptos esenciales. Panamericana</li> <li>- Lehninger, Nelson y Cox (2006). Principios de Bioquímica. Omega</li> <li>- Stryer, Berg y Tymoczko (2009). Bioquímica 6ª Edn. Reverte</li> <li>- Tymoczko, Berg, Stryer (2014). Bioquímica curso básico. Reverté</li> </ul> <p>No moodle da asignatura incluíranse ligazóns a páxinas web e outras fontes bibliográficas.</p>
Complementary	- Melo y Cuamatzi (2004). Bioquímica de los procesos metabólicos. Reverté-UAM Xochimilco



## Recommendations

### Subjects that it is recommended to have taken before

Química/610G02001

Bioquímica: Bioquímica I/610G02011

### Subjects that are recommended to be taken simultaneously

### Subjects that continue the syllabus

Bioquímica e Bioloxía Molecular/610G02013

Fundamentos bioquímicos de biotecnoloxía/610G02014

### Other comments

We inform that to be honored with "matrícula" the best qualified students in the first opportunity (January) will have preference.

Drinks or food are not allowed in the classroom. We recommend the assistance to the reduced groups and the personal tutorials to increase the student's success.

(\* )The teaching guide is the document in which the URV publishes the information about all its courses. It is a public document and cannot be modified. Only in exceptional cases can it be revised by the competent agent or duly revised so that it is in line with current legislation.