

		Teaching Guide			
	Identifyi	ng Data			2018/19
Subject (*)	Molecular Techniques			Code	610441002
Study programme	Mestrado Universitario en Bioloxía Molecular, Celular e Xenética				
		Descriptors			
Cycle	Period	Year		Туре	Credits
Official Master's Degre	ee 1st four-month period	First		Obligatory	6
Language	SpanishGalicianEnglish				
Teaching method	Face-to-face				
Prerequisites					
Department	Bioloxía				
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Web	ciencias.udc.es/masters-bcm/master-en-biología-molecular-y-celula				
General description					

Codo	Study programme competences / results
Code	Study programme competences / results
A1	Skills of using usual techniques and instruments in the cellular, biological and molecular research: that are able to use techniques and
	instruments as well as understanding potentials of their uses and applications.
A2	Skills of working in a sure way in the laboratories knowing operation handbooks and actions to avoid incidents of risk.
A3	Skills of understanding the functioning of cells through the structural organization, biochemistry, gene expression and genetic variability.
A4	Skills to apply molecular techniques to the study of the plant cell physiology, its response to external triggers and their biotechnological
	applications.
A5	Skills of understanding the microorganisms' role as pathogenic agents and as biotechnological tools.
A8	Skills of having an integrated view of the previously acquired knowledge about Molecular and Cellular Biology and Genetics, with an
	interdisciplinary approach and experimental work.
A9	Skills of understanding the structure and dynamics of proteins to individual and proteomic level, as well as the techniques that are
	necessary to analyze them and to study their interactions with other biomolecules.
A10	Skills of modifying genes, proteins and chromosomes with biotechnological applications
A12	Skills to understand, detect and analyze the genetic variation, knowing genotoxicity processes and methodologies for its evaluation, as
	well as carrying out diagnosis and genetic risk studies.
A13	Skills to become a professional in health, pharmacy, veterinary, animal production, biotechnology or food sectors.
B1	Analysis skills to understand biological problems in connection with the Molecular and Cellular Biology and Genetics.
B2	Skills of decision making for the problem solving: that are able to apply theoretical knowledges and practical acquired in the formulation
	biological problems and the looking for solutions.
B3	Skills of management of the information: that are able to gather and to understand relevant information and results, obtaining conclusion
	and to prepare reasoned reports on scientific and biotechnological questions
B4	Organization and work planning skills: that are able to manage the use of the time as well as available resources and to organize the wo
	in the laboratory.
C6	Considering critically the knowledge, technologies and the available information to solve problems with which should face.

Learning outcomes	
Learning outcomes	Study programme
	competences /
	results



Handle the necessary equipment for cellular and molecular techniques.	AR1		
	AR2		
Know the protocols used for the different techniques.	AR1		
	AR2		
Know the applications for the different techniques.	AR1	BR2	CC6
	AR4		
	AR5		
	AR13		
Consider the ways to resolve the methodological problems associated with the performance of the techniques.		BR1	
Establish the relationships between the different techniques used and its possible combination to resolve the problems.		BR1	
Interpret data from observations and measurements in the laboratory.		BR3	
Plan, design and conduct experiments related with the techniques learned.		BR2	
		BR4	
Maintain a critical attitude for a perfect experimental work.			CC6
Relate the chemical and structural properties of biomolecules with laboratory techniques that are most suitable for isolation,	AR1	BR1	
purification and characterization.	AR9	BR2	
Know in depth the possibilities and characteristics of PCR and real-time PCR.	AR2	BR3	
		BR4	
Understand and handle the techniques of recombinant DNA that can be used for analysis and manipulation of biomolecules.	AR1	BR2	
	AR2		
	AR8		
	AR10		
Use methods and techniques to detect and analyze genetic variation.	AR1	BR3	
	AR3		
	AR12		

Contents			
Торіс	Sub-topic		
Purification of Biomolecules	Principle of centrifugation technique and instrumentation. Preparative and Analytical		
	Centrifugation.		
	Chromatographic Techniques: principle and selection criteria.		
	Electrophoresis: principle and types. Isoelectric focusing technique. Capillary		
	electrophoresis.		
PCR	Advanced concepts in PCR		
	Differences between PCR and Real-time PCR		
	Detection methods of amplicons		
	Trial design and results analysis		
Tecnology of molecular markers	Concept and single nucleotide polymorphisms (SNPs)		
	Protein markers		
	DNA markers based in Nucleic Acid Hibridization		
	Pattern Multi Locus by PCR techniques		
	DNA markers based in PCR Mono-locus		
	Single nucleotide polymorphisms (SNPs)		



Recombinant DNA	Enzymes and protocols used in recombinant DNA techniques
	Genomics GeneBank
	Expression GeneBank
	GeneBank analysis
	Transfer and Blotting techniques
	Sequencing techniques
	Site-direct mutagenesis techniques
	Silencing techniques
	Transgenic organisms: uses and applications

	Plannin	g		
Methodologies / tests	Competencies /	Teaching hours	Student?s personal	Total hours
	Results	(in-person & virtual)	work hours	
Guest lecture / keynote speech	A1 A4 A5 A10 A13	14	14	28
Laboratory practice	A1 A2 A3 A12 B4	24	48	72
Supervised projects	A1 A3 A8 A9 B1 B3	0	42	42
	B2			
Mixed objective/subjective test	A1 A3 A9 A12 B1 B2	2	4	6
	C6			
Personalized attention		2	0	2
(*)The information in the planning table is for	r guidance only and does not	take into account the l	neterogeneity of the stud	lents.

	Methodologies		
Methodologies	Description		
Guest lecture /	By the Professors or/and by the exhibition of student work		
keynote speech			
Laboratory practice	Practical classes in the laboratory; Problems solving and practical cases		
Supervised projects	Research Project related with the techniques made in the laboratory. It will be develop individually under Professor?s supervisión.		
Mixed	Exam about theoretical and practical subjects.		
objective/subjective			
test			

	Personalized attention
Methodologies	Description
Supervised projects	Personalized tutoring focused on guidance to help the students: resolving doubts and clarifications.
Guest lecture /	
keynote speech	The tutoring schedule will be indicated the first class by each Professor. The students may request an appointment and/or
Laboratory practice	resolving doubts by e-mail.

	Assessment			
Methodologies	Competencies /	Description Qualific		
	Results			
Supervised projects	A1 A3 A8 A9 B1 B3	Elaboration and writing of a supervised work.	30	
	B2			
Laboratory practice	A1 A2 A3 A12 B4	Along the practical classes, the students will answer questions and problems, which	20	
		will be part of the continuous evaluation of the course.		



Mixed	A1 A3 A9 A12 B1 B2	Exam with questions in which the student must apply the knowledge and skills	50
objective/subjective	C6	acquired along the course.	
test			

Assessment comments

.-The evaluation criteria listed will be apply to two types of registration (classroom and blended learning).

.-The attendance to Practical clases is a necessary condition to be evaluated.

.-The qualifications obtained with the Supervised Project and Practical Exercises will be maintained for the 2° Option (July) if the student do not pass the Final Exam in the 1° Option (January), and in the Final Qualification Records (QRs) will appear the qualification of 4.

.-According to the rule of qualifications and records in Grades and Masters, the Quality Committee of the Faculty of Sciences, agreed to the recommendation to concede the ?Honors Qualification? to those students who obtained the highest marks in the 1st Option\_June.

	Sources of information
Basic	- M. L. Marina, A. Ríos, M. Valcárcel (2005). Analysis and detection by capillary electrophoresis . Amsterdam :
	Elsevier
	- Westermeier, Reiner. (2005). Electrophoresis in practice : a guide to methods and applications of DNA and protein
	separations. Weinheim : Wiley-VCH
	- Weiner MP, Gabriel SB, Stephens JC, (2007). Genetic variation: a laboratory manual. Cold Spring harbor Laborator
	Press, New York.
	- Brown TA (2008). Genomes (3º ed) Médica Panamericana, Buenos Aires.
	- Morteza G. Khaledi (1998). High-performance capillary electrophoresis theory, techniques, and applications . New
	York : John Wiley & Sons,
	- Nuez F, Carrillo JM, (2000). Los marcadores genéticos en la mejora vegetal Universidad Politécnica de Valencia.
	- Avise CJ (2004). Molecular markers, natural history, and evolution (2ª ed.) Sinauer Associates, Sunderland, MA.
	- Keith Wilson and John Walker (1995). Principles and Techniques of Practical Biochemistry. Cambridge, University
	Press
	- Dorak, T. (2007). Real-Time PCR. Routledge Taylor and Francis.
	- Mackay, I. M. (2007). Real-time PCR in microbiology : from diagnosis to characterisation. Norfolk: Caister Academic
	Press.
	- Edwards, K., Logan J. & amp; Saunders, N. (2004). Real-time PCR: an essential guide Horizon bioscience.
	- Logan J, Edawards K, Saunders N. (2009). Real-Time PCR: Current Technology and applications Caister
	Academic Press
Complementary	In addition, it will be provide research articles on the topics covered during the course in the Moodle virtual platform.Ir
	addition, it will be provide research articles on the topics covered during the course in the Moodle virtual platform.

Recommendations
Subjects that it is recommended to have taken before
Subjects that are recommended to be taken simultaneously
Subjects that continue the syllabus
Other comments



(\*)The teaching guide is the document in which the URV publishes the information about all its courses. It is a public document and cannot be modified. Only in exceptional cases can it be revised by the competent agent or duly revised so that it is in line with current legislation.