



Teaching Guide				
Identifying Data			2022/23	
Subject (*)	Cellular Techniques		Code	610441001s
Study programme	Máster Universitario en Bioloxía Molecular, Celular e Xenética (semipresencial)			
Descriptors				
Cycle	Period	Year	Type	Credits
Official Master's Degree	1st four-month period	First	Obligatory	6
Language	SpanishGalicianEnglish			
Teaching method	Hybrid			
Prerequisites				
Department	BioloxíaDepartamento profesorado máster			
Coordinador	Yañez Sanchez, Julian	E-mail	julian.yanez@udc.es	
Lecturers	Bernal Pita da Veiga, María de los Ángeles Castro Castro, Antonio Manuel Insua Pombo, Ana María López Armada, María José Rioboo Blanco, Carmen Yañez Sanchez, Julian	E-mail	angeles.bernal@udc.es antonio.castro@udc.es ana.insua@udc.es maria.jose.lopez.armada@col.udc.es carmen.rioboo@udc.es julian.yanez@udc.es	
Web	campusvirtual.udc.gal/login/index.php			
General description	Experimental subject focused on microscopy (including image analysis), plant and animal cell culture, flow cytometry and cytogenetic techniques.			

Study programme competences / results	
Code	Study programme competences / results
A1	Skills of working in a sure way in the laboratories knowing operation handbooks and actions to avoid incidents of risk.
A2	Skills of using usual techniques and instruments in the cellular, biological and molecular research: that are able to use techniques and instruments as well as understanding potentials of their uses and applications.
A13	Skills to become a professional in health, pharmacy, veterinary, animal production, biotechnology or food sectors.
B3	Skills of management of the information: that are able to gather and to understand relevant information and results, obtaining conclusions and to prepare reasoned reports on scientific and biotechnological questions
B4	Organization and work planning skills: that are able to manage the use of the time as well as available resources and to organize the work in the laboratory.

Learning outcomes			
Learning outcomes		Study programme competences / results	
To understand the theoretical foundations on which microscopy (including image analysis), (plant and animal) cell culture, flow cytometry and cytogenetic techniques are based.	AR1		
	AR2		
To acquire basic skills in the management and use of instrumental and units required for the development of cellular techniques.	AR1		
	AR2		
	AR13		
To know the applications of the different cellular techniques.	AR2		
To design, plan and conduct experiments regarding the techniques learned.	AR1	BR3	
	AR2	BR4	
Adequate oral and written expression in the official languages.			
Using ICT in working contexts and lifelong learning.			
Ability to manage times and resources: developing plans, prioritizing activities, identifying critical points, establishing goals and accomplishing them.			



Contents	
Topic	Sub-topic
Microscopy and image analysis	Fundamentals, techniques and applications of light and electron microscopy. Fluorescence and confocal scanning microscopy: advanced techniques and applications. Introduction to image processing and image analysis
Cellular cultures	Introduction to cell cultures. Types of cell cultures. Cell culture requirements. Quantification of cellular parameters. Contamination. Cytotoxicity. In vitro cultures of plant tissues. Callus. Cultivation of plant cells in suspension.
Flow cytometry	General principles and methods of cytometry. Sample preparation and standardization of analysis procedures. Functional analysis of cells.
Cytogenetic techniques	Chromosome preparation and karyotype. Conventional in situ hybridization. Advanced techniques of fluorescence in situ hybridization (FISH).

Planning				
Methodologies / tests	Competencies / Results	Teaching hours (in-person & virtual)	Student's personal work hours	Total hours
Document analysis	A2	0	40	40
Laboratory practice	A1 A2 A13 B3 B4	28	42	70
Supervised projects	A2 B3 B4 C1 C3 C8 C9	0	19	19
Mixed objective/subjective test	A2 B3	2	15	17
Personalized attention		4	0	4

(*)The information in the planning table is for guidance only and does not take into account the heterogeneity of the students.

Methodologies	
Methodologies	Description
Document analysis	This methodology involves watching videos of lectures, reading documents prepared by the lecturers and/or reading bibliographic documents on the theoretical-practical fundamentals and aspects related to the different techniques used in Cell Biology.
Laboratory practice	Practical sessions represent an indispensable part of the course, in which practical aspects and applications of the different cellular techniques will be approached. Practical sessions will be developed in concrete laboratories and places: laboratories of the Faculty of Sciences, Laboratories of Biomedical Research Institute (INIBIC), Scientific Research Support Services (SAI) of UDC. Students will develop laboratory protocols and attend demonstrations about the use of research units.
Supervised projects	Students must carry out works, resolve problems and/or questions about specific aspects of the techniques used.
Mixed objective/subjective test	It will consist of a written exam with questions-test and/or short answer questions about theoretical and practical contents and applications of the cellular techniques.

Personalized attention	
Methodologies	Description
Supervised projects Document analysis	Students (individually or in small groups) may consult their doubts about the contents and activities of the subject via phone and/or electronic support. A forum on the Virtual Campus/Teams may be used for the formulation of doubts/comments.

Assessment			
Methodologies	Competencies / Results	Description	Qualification



Supervised projects	A2 B3 B4 C1 C3 C8 C9	The ability to design (and plan) experiments, interpretate data and solve problems will be assessed.	30
Mixed objective/subjective test	A2 B3	The acquisition of knowledge about the theoretical foundations and applications, clarity of explanations, ability to integrate and link information handled and the ability to interpret data and solve problems will be taken into account.	50
Laboratory practice	A1 A2 A13 B3 B4	Attendance, skill in scheduled tasks and knowledge of the potential risks in laboratory practices will be assessed.	20

Assessment comments

The exam will take place through the "Virtual Campus" on the dates specified in the official calendar.

Attendance at the practical

lessons is a necessary condition to be evaluated. In case of not passing the evaluation in the first opportunity of the call, the grades obtained in the supervised works and practices will be kept for the second opportunity. Honors will be awarded preferably among students (face-to-face and non-face-to-face) presented in the evaluation corresponding to the first opportunity of the call.

Fraudulent performance of the tests or

assessment activities, once verified, will directly imply the failing grade

"0" in the call.

Sources of information



Basic	CULTIVOS CELULARES Basra, A.S. (2000). Plant growth regulators in agriculture and horticulture. Their role and commercial uses. Ed. Food Products Press. Benítez Burraco, A. (2005). Avances recientes en Biotecnología vegetal e ingeniería genética de plantas. Editorial Reverté. Boulton, A.A. e col. (1992). Practical cell culture techniques. Humana Press. Butler, M. (2008). Animal cell culture and technology. Taylor & Francis, 2nd edition. Collin, H.A. e Edwards, S. (1998). Plant cell culture. Guilford Bios Scientific Publishers. Davis, J.M. (2011). Animal cell culture. Essential methods. Wiley-Blackwell. Doyle, A. e Griffiths, J.B. (2000). Cell and tissue culture for medical research. John Wiley and Sons. Fedoroff, S. e Richardson, A. (1992). Protocols for neural cell culture. Humana Press. Freshney, R.I. (2010). Culture of animal cells. A manual of basic technique and specialized applications. Wiley-Liss, Inc. 6ª Edition. Hammond, J., McGarvey, P., Yusibov, V. (1999). Plant Biotechnology. New products and Applications. Springer Verlag. Loyola-Vargas, V.M. e Vázquez-Flota F. (2006). Plant cell culture protocols. Humana Press. 2nd Edition. Pollard, J.W. e Walker, J.M. (1997). Basic cell culture protocols. Humana Press. Shaw, A.J. (1996). Epithelial cell culture. A practical approach. Oxford University Press. Taji, A., Kumar, P., Lakshmanan, P. (2002). In vitro plant breeding. Food Products Press. Trigiano, R.N. e Gray, D.J. (2004). Plant development and biotechnology. CRC Press. Tzfira, T. e Citovsky, V. (2006). Agrobacterium-mediated genetic transformation of plants: biology and biotechnology. Curr. Opin. Biotechnol. 17:147-154. Vunjak-Novakovic, G. & Freshney, R.I. (2006). Culture of cells for tissue engineering. Wiley-Liss, Inc. TÉCNICAS DE MICROSCOPIA E ANÁLISE DE IMAGEM Watt, Ian M. (1996). The principles and practice of electron microscopy. Cambridge University Press. Hoppert, M. (1998). Electron microscopy in microbiology. Bios Scientific Publishers. Bozzola, John J. (1999). Electron microscopy : principles and techniques for biologists. Jones and Bartlett Publishers. Dykstra, Michael J. (2003). Biological electron microscopy theory, techniques, and troubleshooting. Kluwer Academic/Plenum Publishers. Robin Harris. (1991). Electron microscopy in biology a practical approach. Oxford University Press. Hunter, Elaine Evelyn. (1984). Practical electron microscopy a beginner's illustrated guide. Praeger, cop. Slayter, Elizabeth M. (2000). Light and electron microscopy. Cambridge University Press. Herman, B. (1998). Fluorescence microscopy. Bios Scientific Publishers. Donat-P. Häder. (1992). Image analysis in biology. CRC Press, cop. Pertusa, JF. (2003). Técnicas de Análisis de imagen. Aplicaciones en Biología. Publicaciones de la Universidad de Valencia. CITOMETRÍA DE FLUJO Omerod, M.G. (2009). Flow Cytometry: A Basic Introduction. 2a Ed. IRL Practical Approach series. Oxford University Press. Shapiro, H.M. (2004). Practical flow cytometry. Wiley-Liss. 4a ed. New York. TÉCNICAS CITOXENÉTICAS Czepulkowski, B. (2001). Analyzing chromosomes. BIOS Scientific Publishers, Oxford. Gersen, S.L., Keagle, M.B. (2013). The principles of clinical cytogenetics. Springer, New York. Gosden, J.R. (1994). Chromosome analysis protocols. Humana Press, Totowa (New Jersey). Kianian, S.F., Kianian P.M.A. (2016). Plant cytogenetics: methods and protocols. Springer, New York. Liehr, T. (2006). Multicolor FISH in human cytogenetics. Karger, Basel. Liehr, T. (2009). Fluorescence in situ hybridization (FISH)-application guide. Springer-Verlag, Berlin. Leitch, A.R., Schwarzacher, T., Jackson, D. (1994). In situ hybridization: a practical guide. Bios Scientific Publishers, Oxford. Verma, R.S. e Babu, A. (1989). Human chromosomes: manual of basic techniques. Pergamon Press, New York.
Complementary	- Artigos científicos sobre temas relacionados coa materia proporcionados a través da plataforma Moodle. - Páxinas web Xeral PubMed: http://www.ncbi.nlm.nih.gov/pubmed Cultivos Celulares-https://inmunomundo.files.wordpress.com/2015/12/cultivo-celular.pdf http://www.lgcstandards-atcc.org/Cito metría-Cytometry: http://www3.interscience.wiley.com/cgi-bin/jhome/33945 Microscopía e Análise de imaxe http://zeiss-campus.magnet.fsu.edu/index.html http://www.microscopyu.com/tutorials http://www.olympusfluoview.com/index.html http://w3.uniroma1.it/MEDICFISIO/microscopy.htm http://rsbweb.nih.gov/ij/index.html http://www.invitrogen.com/site/us/en/home/support/Research-Tools/Fluorescence-SpectraViewer.html

Recommendations

Subjects that it is recommended to have taken before

Subjects that are recommended to be taken simultaneously



Subjects that continue the syllabus

Other comments

Program Green Campus

Empower of SciencesTo help to achieve some sustainable immediate surroundings

and fulfil with the point 6 of the Environmental Statement of the faculty of

Sciences (2020), the documentary works that realise in this matter:to. They will request

mostly in virtual format and computer supportb. To realise in

paper:-they will not employ

plastic-they will realise

impressions to double expensive-they will employ paper

recycled-they will avoid the

realisation of draftsTo Environmental Statement is available

in:https://ciencias.udc.es/images/Facultade/Green_Campus/Regulamento_Comit%C3%A9_Green_Campus_FCiencias.pdf

(*)The teaching guide is the document in which the URV publishes the information about all its courses. It is a public document and cannot be modified. Only in exceptional cases can it be revised by the competent agent or duly revised so that it is in line with current legislation.