		Teaching Gui	ide		
	Identifying Data	a			2023/24
Subject (*)	Cellular Techniques		Code	610441001s	
Study programme	Máster Universitario en Bioloxía Molecular, Celular e Xenética (semipresencial)			'	
		Descriptors			
Cycle	Period Year Ty		Туре	Credits	
Official Master's Degre	e 1st four-month period	First		Obligatory	6
Language	SpanishGalicianEnglish				·
Teaching method	Hybrid				
Prerequisites					
Department	BioloxíaDepartamento profesorado máster				
Coordinador	Insua Pombo, Ana Maria E-mail ana.insua@udc.es				.es
Lecturers	Bernal Pita da Veiga, María de los Ánge	eles	E-mail	angeles.bernal@	@udc.es
	Castro Castro, Antonio Manuel			antonio.castro@	udc.es
	Folgueira Otero, Mónica			m.folgueira@ud	lc.es
	Insua Pombo, Ana Maria			ana.insua@udc	.es
	Rioboo Blanco, Carmen			carmen.rioboo@	@udc.es
	Yañez Sanchez, Julian			julian.yanez@u	dc.es
Web	campusvirtual.udc.gal/login/index.php				
General description	Experimental subject focused on micros	scopy (includin	g image analy	sis), plant and anima	I cell culture, flow cytometry a
	cytogenetic techniques.				

	Study programme competences
Code	Study programme competences
A1	Skills of working in a sure way in the laboratories knowing operation handbooks and actions to avoid incidents of risk.
A2	Skills of using usual techniques and instruments in the cellular, biological and molecular research: that are able to use techniques and
	instruments as well as understanding potentials of their uses and applications.
A13	Skills to become a professional in health, pharmacy, veterinary, animal production, biotechnology or food sectors.
В3	Skills of management of the information: that are able to gather and to understand relevant information and results, obtaining conclusions
	and to prepare reasoned reports on scientific and biotechnological questions
B4	Organization and work planning skills: that are able to manage the use of the time as well as available resources and to organize the work
	in the laboratory.
C1	Ability to express oneself correctly, both orally and in writing, in the official languages of the autonomous community
С3	Using ICT in working contexts and lifelong learning.
C8	Valuing the importance of research, innovation and technological development for the socioeconomic and cultural progress of society.
C9	Ability to manage times and resources: developing plans, prioritizing activities, identifying critical points, establishing goals and
	accomplishing them.

Learning outcomes				
Learning outcomes			Study programme	
		competences		
To understand the theoretical foundations on which microscopy (including image analysis), (plant and animal) cell culture, flow	AR1		CC1	
cytometry and cytogenetic techniques are based.	AR2		CC8	
To acquire basic skills in the management and use of instrumental and units required for the development of cellular			CC3	
techniques.	AR2			
	AR13			
To know the applications of the different cellular techniques.	AR2		CC1	
			CC8	
To design, plan and conduct experiments regarding the techniques learned.	AR1	BR3	CC3	
	AR2	BR4	CC9	

Contents				
Topic	Sub-topic			
Microscopy and image analysis	Fundamentals, techniques and applications of light and electron microscopy.			
	Fluorescence and confocal scanning microscopy: advanced techniques and			
	applications. Introduction to image processing and image analysis			
Cellular cultures	Introduction to cell cultures. Types of cell cultures. Cell culture requirements.			
	Quantification of cellular parameters. Contamination. Cytotoxicity.			
	In vitro cultures of plant tissues. Callus. Cultivation of plant cells in suspension.			
Flow cytometry	General principles and methods of cytometry. Sample preparation and standardization			
	of analysis procedures. Functional analysis of cells.			
Cytogenetic techniques	Chromosome preparations. Chromosome staining and banding. Standard			
	fluorescence in situ hybridizationn (FISH). Advanced techniques of (FISH).			

Planning			
Competencies	Ordinary class	Student?s personal	Total hours
	hours	work hours	
A2	0	40	40
A1 A2 A13 B3 B4	28	42	70
A2 B3 B4 C1 C3 C8	0	19	19
C9			
A2 B3	2	15	17
	4	0	4
	A2 A1 A2 A13 B3 B4 A2 B3 B4 C1 C3 C8 C9	A2 0 A1 A2 A13 B3 B4 28 A2 B3 B4 C1 C3 C8 0 C9 A2 B3	Competencies         Ordinary class hours         Student?s personal work hours           A2         0         40           A1 A2 A13 B3 B4         28         42           A2 B3 B4 C1 C3 C8         0         19           C9         A2 B3         2         15

Methodologies				
Methodologies	gies Description			
Document analysis	This methodology involves watching videos of lectures, reading documents prepared by the lecturers and/or reading			
	bibliographic documents on the theoretical-practical fundamentals and aspects related to the different techniques used in Cell			
	Biology.			
Laboratory practice	Practical sessions represent an indispensable part of the course, in which practical aspects and applications of the different			
	cellular techniques will be approached. Practical sessions will be developed in concrete laboratories and places: laboratories			
	of the Faculty of Sciences, Laboratories of Biomedical Research Institute (INIBIC), Scientific Research Support Services (SAI)			
	of UDC.			
	Students will develop laboratory protocols and attend demonstrations about the use of research units.			
Supervised projects	Students must carry out works, resolve problems and/or questions about specific aspects of the techniques used.			
Mixed	It will consist of a written exam with questions-test and/or short answer questions about theoretical and practical contents and			
objective/subjective	applications of the cellular techniques.			
test				

Personalized attention				
Methodologies	Methodologies Description			
Supervised projects	Students (individually or in small groups) may consult their doubts about the contents and activities of the subject via phone			
Document analysis	and/or electronic support.			
	A forum on the Virtual Campus/Teams may be used for the formulation of doubts/comments.			

Assessment					
Methodologies	Competencies	Description	Qualification		

Supervised projects	A2 B3 B4 C1 C3 C8	The ability to design (and plan) experiments, interpretate data and solve problems will	30
	C9	be assessed.	
Mixed	A2 B3	The acquisition of knowledge about the theoretical foundations and applications,	50
objective/subjective		clarity of explanations, ability to integrate and link information handled and the ability	
test		to interpret data and solve problems will be taken into account.	
Laboratory practice	A1 A2 A13 B3 B4	Attendance, skill in scheduled tasks and knowledge of the potential risks in laboratory	20
		practices will be assessed.	

## **Assessment comments**

The exam will take place through the "Virtual Campus" on the dates specified in the official calendar.

Attendance at the practical

lessons is a necessary condition to be evaluated.

In case of not passing the

evaluation in the first opportunity of the call, the grades obtained in the

supervised works and practices will be kept for the second opportunity.

Honors

will be awarded preferably among students (face-to-face and non-face-to-face)

presented in the evaluation corresponding to the first opportunity of the call.

In case of fraud in tests or evaluation activities, the regulations in force at UDC will be applied.

Sources of information



#### Basic

CULTIVOS CELULARESBasra, A.S. (2000). Plant growth regulators in agriculture and horticulture. Their role and commercial uses. Ed. Food Products Press. Benítez Burraco, A. (2005). Avances recientes en Biotecnología vegetal e ingeniería genética de plantas. Editorial Reverté.Boulton, A.A. e col. (1992). Practical cell culture techniques. Humana Press.Butler, M. (2008). Animal cell culture and technology. Taylor & Damp; Francis, 2nd edition.Collin, H.A. e Edwars, S. (1998). Plant cell culture. Guilford Bios Scientific Publishers. Davis, J.M. (2011). Animal cell culture. Essential methods. Wiley-Blackwell.Doyle, A. e Griffiths, J.B. (2000). Cell and tissue culture for medical research. John Wiley and Sons. Fedoroff, S. e Richardson, A. (1992). Protocols for neural cell culture. Humana Press. Freshney, R.I. (2010). Culture of animal cells. A manual of basic technique and specialized applications. Wiley-Liss, Inc. 6ª Edition.Hammond, J., McGarvey, P., Yusibov, V. (1999). Plant Biotechnology. New products and Applications. Springer Verlag, Loyola-Vargas, V.M. e Vázguez-Flota F. (2006), Plant cell culture protocols, Humana Press, 2nd Edition. Pollard, J.W. e Walker, J.M. (1997). Basic cell culture protocols. Humana Press. Shaw, A.J. (1996). Epithelial cell culture. A practical approach. Oxford University Press. Taji, A., Kumar, P., Lakshmanan, P. (2002). In vitro plant breeeding. Food Products Press.Trigiano, R.N. e Gray, D.J. (2004). Plant development and biotechnology. CRC Press.Tzfira, T. e Citovsky, V. (2006). Agrobacterium-mediated genetic transformation of plants: biology and biotechnology. Curr. Opin. Biotechnol. 17:147?154. Vunjak-Novakovic, G.& Freshney, R.I. (2006). Culture of cells for tissue engineering. Wiley-Liss, Inc.TÉCNICAS DE MICROSCOPÍA E ANÁLISE DE IMAXEWatt, Ian M. (1996). The principles and practice of electron microscopy. Cambridge University PressHoppert, M. (1998). Electron microscopy in microbiology. Bios Scientific PublishersBozzola, John J. (1999). Electron microscopy: principles and techniques for biologists. Jones and Bartlett Publishers. Dykstra, Michael J. (2003). Biological electron microscopy theory, techniques, and troubleshooting. Kluwer Academic/Plenum PublishersRobin Harris. (1991). Electron microscopy in biology a practical approach. Oxford University Press.Hunter, Elaine Evelyn. (1984). Practical electron microscopy a beginner's illustrated guide. Praeger, cop.Slayter, Elizabeth M. (2000). Light and electron microscopy. Cambridge University PressHerman, B. (1998). Fluorescence microscopy. Bios Scientific PublishersDonat-P. Häder. (1992). Image analysis in biology. CRC Press, cop. Pertusa, JF. (2003). Técnicas de Análisis de imagen. Aplicaciones en Biología. Publicaciones de la Universidad de Valencia. CITOMETRÍA DE FLUXOOrmerod, M.G. (2009). Flow Cytometry: A Basic Introduction. 2a Ed. IRL Practical Approach series. Oxford University Press.Shapiro, H.M. (2004). Practical flow cytometry. Wiley-Liss. 4a ed. New York.TÉCNICAS CITOXENÉTICASCzepulkowski, B. (2001). Analyzing chromosomes. BIOS Scientific Publishers, Oxford.Gersen, S.L., Keagle, M.B. (2013). The principles of clinical cytogenetics. Springer, New York.Gosden, J.R. (1994). Chromosome analysis protocols. Humana Press, Totowa (New Jersey), Kianian, S.F., Kianian P.M.A. (2016). Plant cytogenetics: methods and protocols. Spinger, New YorlLiehr, T. (2006). Multicolor FISH in human cytogenetics. Karger, Basel.Liehr, T. (2009). Fluorescence in situ hybridization (FISH)-application guide. Springer-Verlag, Berlin.Leitch, A.R., Schwarzacher, T., Jackson, D. (1994). In situ hybridization: a practical guide. Bios Scientific Publishers, Oxford. Verma, R.S. e Babu, A. (1989). Human chromosomes: manual of basic techniques. Pergamon Press, New York.

### Complementary

- Artigos científicos sobre temas relacionados coa materia proporcionados a través da plataforma Moodle . Páxinas web Xeral PubMed: http://www.ncbi.nlm.nih.gov/pubmed Cultivos Celulares
- -https://inmunomundo.files.wordpress.com/2015/12/cultivo-celular.pdf-http://www.lgcstandards-atcc.org/-https://www.ibiantech.com/el-ayer-y-hoy-de-las-tecnicas-y-medios-de-cultivo-celular/
- -https://www.thermofisher.com/es/es/home/references/gibco-cell-culture-basics/introduction-to-cell-culture.html
  Citometría -Cytometry: http://www3.interscience.wiley.com/cgi-bin/jhome/33945 Microscopía e Análise de imaxe
  http://zeiss-campus.magnet.fsu.edu/index.htmlhttp://www.microscopyu.com/tutorials/http://www.olympusfluoview.com/i
  ndex.htmlhttp://w3.uniroma1.it/MEDICFISIO/microscopy.htmhttp://rsbweb.nih.gov/ij/index.htmlhttp://www.invitrogen.co
  m/site/us/en/home/support/Research-Tools/Fluorescence-SpectraViewer.html

# Recommendations

Subjects that it is recommended to have taken before



Subjects that are recommended to be taken simultaneously

Subjects that continue the syllabus

### Other comments

Gender perspective in this subject, the gender perspective will be taken into account, sexist attitudes will not be tolerated and the values of respect and equality will be promoted. Program Green Campus

Empower of SciencesTo help to achieve some sustainable immediate surroundings

and fulfil with the point 6 of the Environmental Statement of the faculty of

Sciences (2020), the documentary works that realise in this matter:a. They will request mostly in virtual format and computer supportb. To realise in paper:-they will not employ plastic-they will realise impressions to double expensive-they will employ paper

recycled-they will avoid the

realisation of draftsTo Environmental Statement is available

in:https://ciencias.udc.es/images/Facultade/Green\_Campus/Regulamento\_Comit%C3%A9\_Green\_Campus\_FCiencias.pdf

(\*)The teaching guide is the document in which the URV publishes the information about all its courses. It is a public document and cannot be modified. Only in exceptional cases can it be revised by the competent agent or duly revised so that it is in line with current legislation.