Teaching Guide				
Identifying Data				2015/16
Subject (*)	Xenética molecular		Code	610G02020
Study programme	Grao en Bioloxía			
Descriptors				
Cycle	Period	Year	Type	Credits
Graduate	1st four-month period	Third	Obligatoria	6
Language	Galician			
Teaching method	Face-to-face			
Prerequisites				
Department	Bioloxía Celular e Molecular			
Coordinador	Insua Pombo, Ana Maria	E-mail	ana.insua@udc.es	
Lecturers	Insua Pombo, Ana Maria Nantón Varela, Ana Torrecilla Pérez, Zeltia	E-mail	ana.insua@udc.es ana.nanton@udc.es zeltia.torrecilla@udc.es	
Web				
General description	This course focuses on the conceptual and methodological bases necessary to understand the organization, expression, variation and manipulation of genetic material. It provides a molecular perspective to knowledge in "Genetics" (second year) and knowledge necessary to address "Population Genetics and Evolution", "Cytogenetics" and related courses of third and fourth year.			

Study programme competences	
Code	Study programme competences
A5	Analizar e caracterizar mostras de orixe humana.
A11	Identificar e analizar material de orixe biolóxica e as súas anomalías.
A12	Manipular material xenético, realizar análises xenéticas e levar a cabo asesoramento xenético.
A15	Deseñar e aplicar procesos biotecnológicos.
A29	Impartir coñecementos de Bioloxía.
A30	Manexar adecuadamente instrumentación científica.
A31	Desenvolverse con seguridade nun laboratorio.
B1	Aprender a aprender.
B2	Resolver problemas de forma efectiva.
B3	Aplicar un pensamento crítico, lóxico e creativo.
B5	Traballar en colaboración.
B7	Comunicarse de maneira efectiva nunha contorna de traballo.

Learning outcomes			
Learning outcomes	Study programme competences		
General knowledge and understanding of the molecular basis of the organization, expression, variation and manipulation of genetic material	A11 A12 A15 A29	B1 B2 B3 B5 B7	

Knowledge of the basic methodologies used in Molecular Genetics.	A5 A11 A12 A15 A29 A30 A31	B1 B2 B3 B5	
Ability to use sources of information of interest in Molecular Genetics.	A5 A11 A12 A15 A29	B1 B2 B3	
Ability to interpret and transmit information of Molecular Genetics	A29	B1 B2 B3 B5 B7	

Contents	
Topic	**Sub-topic**
1.- GENOME ORGANIZATION	C-value paradox. Prokaryotic and eukaryotic genomes. Single-copy and repetitive DNA sequences. Gene families. Centromeres. Telomeres. Organelle genomes.
2.- DNA REPLICATION	Semiconservative DNA replication: the Meselson and Stahl experiment. Modes of replication. Enzymology of the replication. DNA replication in Escherichia coli. DNA replication in eukaryotes. Telomere synthesis. Replication of mithocondrial and cloroplast DNA.
3.- SYNTHESIS AND PROCESSING OF RNA	Clases of RNA. RNA polymerases. Promoters and transcriptional appartatus. Transcription in prokaryotes and eukaryotes: initiation, elongation and termination. Interrupted genes: exons and introns. Processing of eukaryotic pre-mRNA. Synthesis and processing of pre-rRNA. Synthesis and processing of pre-tRNA. RNA edition. Revision of gene concept.
4.- TRANSLATION	The one gene-one enzyme hypothesis. The genetic code: characteristics and experiments to decipher the code. Initiation of translation. Elongation of the polypeptide chain. Termination of translation. Messenger RNA surveillance.
5.- MUTATION AND DNA REPAIR	Molecular basis of spontaneous mutations: replication errors, unequal crossing over, spontaneous chemical changes. Molecular basis of induced mutations: chemical and physical agents. Repair mechanisms: direct reversal of damaged DNA, excision repair, postreplication repair, error-prone repair, repair of double-strand breaks.
6.- MOLECULAR MECHANISM OF GENETIC RECOMBINATION	The role of genetic recombination. Gene conversion. Models of homologous recombination: Holliday model and double-strand break model. Enzymes required for recombination. Site-specific recombination. Inmunoglobulin genes assemble by recombination.
7.- TRANSPOSABLE GENETIC ELEMENTS	Transposable elements in prokaryotes: insertion sequences, composite transposons and noncomposite transposons. Replicative and non replicative transposition. Transposable elements in eukaryotes: transposons and retrotransposon. Evolutionary significance of transposable elements.
8.- RECOMBINANT DNA TECHNOLOGY	Restriction enzymes. Cloning vectors. DNA libraries: construction and screening. Southern and northern blotting. Restriction maps. Sequencing. PCR. Site-directed mutagenesis.

9.- APPLICATIONS OF RECOMBINANT DNA TECHNOLOGY	Expression of eukaryotic genes in E. coli. DNA transfer to eukaryotic cells. Transgenic animals. Transgenic plants. Gene therapy. Molecular markers. DNA fingerprinting. Genetic diagnosis. Synthetic genomes.
10.- GENOMICS	Physical and genetic mapping. Whole genome sequencing. Genome annotation. DNA microarrays. Reverse Genetics. Comparative genomics. Metagenomics.
11.- REGULATION OF GENE EXPRESSION IN BACTERIA	Jacob and Monod?s operon model for the regulation of lac genes in E. coli. Positive control of the lac operon. The arabinose operon of E. coli: positive and negative control. The triptophan operon of E. coli: negative control and attenuation. Control by RNA molecules.
12.- REGULATION OF GENE EXPRESSION IN EUKARYOTES	Changes in chromatin structure. DNA metilation. Transcriptional control. RNA processing control. Control of mRNA stability. Control at the level of translation. RNA interference. Epigenetics.
13.- GENETIC CONTROL OF DEVELOPMENT	Basic events of development. Drosophila development stages. Maternal-effect, segmentation and homeotic genes in Drososphila. Homeobox genes in other organisms. General aspects of Caenorhabditis elegans development. Genetic control of flower development in Arabidopsis.
PRACTICE 1: DNA EXTRACTION	DNA extraction from Drosophila melanogaster and human cells. Agarose gel electrophoresis of DNA. DNA quantification.
PRACTICE 2: PCR	PCR amplification of the locus PV92. Analysis of an insertion polymorphism of Alu sequences
PRACTICE 3: DOT-BLOT	Nucleic acids hybridization: detection of microsatellite sequences by dot-blot
PRACTICE 4: BIOINFORMATICS.	Database search and comparison of nucleic acid sequences. Primer design. Identification of ORFs.

Planning				
Methodologies / tests	Competencies	Ordinary class hours	Student?s personal work hours	Total hours
Guest lecture / keynote speech	A5 A11 A12 A15 B2 B3 B7	28	42	70
Seminar	A5 A11 A12 A15 A29 B1 B2 B3 B5 B7	8	20	28
Laboratory practice	A5 A11 A12 A15 A30 A31 B1 B2 B3 B5 B7	15	7.5	22.5
Supervised projects	A5 A11 A12 A15 A29 B1 B2 B3 B5 B7	0	21.5	21.5
Mixed objective/subjective test	A5 A11 A12 A15 A29 B1 B2 B3 B7	6	0	6
Personalized attention		2	0	2
(*)The information in the planning table is for guidance only and does not take into account the heterogeneity of the students.				

Methodologies	
Methodologies	Description
Guest lecture / keynote speech	The teacher explains the main contents of each lesson.
Seminar	Resolution/discussion of questions and problems realted to the subject.
Laboratory practice	The student conducts laboratory experiences following a protocol, under the supervision of the teacher.
Supervised projects	Students resolve questions and problems and/or prepare written documents related to some aspects of the subject. This activity is done in groups.

Mixed objective/subjective test	Objective test, short answer questions and problem resolution.

Personalized attention	
Methodologies	**Description**
Supervised projects	Individually or in group, students can attend tutorial sessions to consult any doubts that might arise from the different activities.

Assessment			
Methodologies	**Competencies**	**Description**	**Qualification**
Supervised projects	A5 A11 A12 A15 A29 B1 B2 B3 B5 B7	Assessment will be based on providing correct answers, clarity of explanations and documentary sources used. The score depends on work carried out at individual (10%) and group level (10%).	20
Mixed objective/subjective test	A5 A11 A12 A15 A29 B1 B2 B3 B7	The degree of general knowledge and understanding of the subject will be assessed. Consists of two parts. One is related to theoretical content and represents 70% of the score. The other is related to the laboraroty practices and represents 10% of the score.	80

Assessment comments
To pass the course, the score must be 5 or higher but with at least a 4 in each part of the test. If the sum of the score of all activities is higher than 5, but the score on one part of the text is lower than 4 then the final score is 4.9 (failing score). It is considered "NP" (non attendance) when less than 30% of the assessed activities were carried out. Preferably, first class honors will be awarded in January among students with a score of 9 or higher. A mid-term exam will be held. A score of 5 or higher will be maintained until July. In July there is the opportunity to retake only the test. The January's score of supervised projects is maintained.

Sources of information	
Basic	- Griffiths, A.J.F., Wessler, S.R., Suzuki, Lewontin, R.C. Carroll, S.B. (2008). Genética. McGraw-Hill/Interamericana de España, Madrid - Klug, W.S., Cummings, M.R., Spencer, C.A (2013). Conceptos de Genética . Pearson/Prentice Hall, Madrid - Pierce, B.A. (2010). Genética: un enfoque conceptual. Médica Panamericana, Madrid

Complementary	- Alberts, B., Johnson, A., Lewis, J., Raff, M., Roberts, K., Walter, P. (2010). Biología Molecular de la célula. Omega, Barcelona - Benito, C., Espino, F.C. (2013). Genética: conceptos esenciales. Médica Panamericana, Madrid - Brooker, R.J. (2005). Genetics: Analysis and Principles (2nd ed). . McGraw-Hill, Boston, USA - Brown, T.A. (2008). Genomas (3ª ed.). Médica Panamericana, Buenos Aires - Hartwell, L.H., Hood, L., Goldberg, M.L., Reynols, A.E., Silver, L.M., Veres, R.C. (2008). Genetics: from genes to genomes (3ª ed.) . McGraw-Hill, Boston, USA - Krebs, J.E., Goldstein, E.S., Kilpatrick, S.T. (2012). Lewin genes: fundamentos. Médica Panamericana, Madrid - Lewin, B. (2008). Genes IX. McGraw-Hill. México - Lodish, H., Berk, A., Zipursky, S.L., Matsudaira, P., Baltimore, D., Darnell, J. (2002). Biología celular y Molecular (4ª ed) . Médica Panamericana, Madrid - Perera, J., Tormo, A., García, J.L. 2002b (2002). Ingeniería genética. Vol. II. Expresión de DNA en sistemas heterólogos. Síntesis, Madrid - Perera, J., Tormo, A., García, J.L. (2002). Ingeniería genética. Vol. I: Preparación, análisis, manipulación y clonaje de DNA. Síntesis, Madrid - Russell, P.J. (2010). iGenetics: a molecular approach (3º ed.) . Benjamin Cummings, San Francisco, USA - Snustad, D.P., Simmons, M.J. (2006). Principles of Genetics (4ed). John Wiley and Sons, Inc. New York, USA Consultar a plataforma Moodle para fontes de información adicionales.

Recommendations
Subjects that it is recommended to have taken before
Citoloxía/610G02007 Bioquímica: Bioquímica I/610G02011 Bioquímica: Bioquímica II/610G02012 Microbioloxía/610G02015 Xenética/610G02019
Subjects that are recommended to be taken simultaneously
Subjects that continue the syllabus
Xenética de poboacións e evolución/610G02021 Citoxenética/610G02022
Other comments
Recommendations: Attend class and follow the development of the course regularly. Check Moodle and email regularly to obtain the materials and know the schedule of activities. Attend tutorials to resolve any questions or difficulties that may arise. Consult the recommended bibliography. Keep up-to-date with course work.